Rapid and Scalable Fabrication of Low Impedance, 3D Dry Electrodes for Physiological Sensing

Ryan Kaveh,* Natalie Tetreault, Karthik Gopalan, Julian Maravilla, Michael Lustig, Rikky Muller, and Ana C. Arias*

1. Introduction

In clinical settings, non-invasive electrophysiological (ExG) recordings such as electrocardiography (ECG), electromyography (EMG), and electroencephalography (EEG) enable expansive brain-machine interfaces,[1] arrhythmia sensors,[2] and advanced prostheses.[3] These devices rely on a low-impedance interface between conductors and skin, achieved by and advanced prostheses. [3] These devices rely on a low-impedance interface between conductors and skin, achieved by

Medical electrophysiological sensors that can study the body and diagnose diseases depend on consistently low impedance electrode–skin interfaces. Clinical-standard wet electrodes use hydrogels and skin abrasion to improve the interface and thus the recorded signal quality. These electrodes are challenging to self-administer and impede in-home care. Wearable dry electrodes are more practical; however, they show higher impedances than wet electrodes and are costly to customize. This work presents a fabrication method for rapidly producing low impedance, anatomically fit dry electrodes that do not require hydrogels. By using electroless copper and gold plating with 3D printing, biocompatible electrodes can be optimized for individuals at a fraction of the cost of existing vacuum deposition-based techniques. Example 3D dry electrodes made with this process are evaluated alongside clinical-standard devices in typical scenarios to compare electrical performance and comfort. The resulting dry electrodes exhibit an average electrode-skin impedance of 66.7 kΩ at 50 Hz and DC offset of −20 mV without any hydrogel, which, when area normalized, is within the range achieved by wet electrodes without skin abrasion.

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extensive skin preparation and application of electrodes that are “wet” with hydrogel. This process typically requires a trained professional but results in recordings with high signal-to-noise ratios (SNR), a requirement for restorative and potentially life-changing treatments. These wet electrode hydrogels ensure consistent and low-impedance electrode-skin contact only until the gel dries out. This drying leads to increased electrode-skin impedance (ESI), which reduces recorded single amplitude and increases susceptibility to power-line interference, ultimately reducing SNR.[4]

The skin preparation required for wet electrodes (especially in EEG setups for sleep/epilepsy studies) also often leads to skin-irritation, hair loss,[5] and discomfort from residual gel left in hair.

Some have attempted to expand these clinical techniques to everyday users by incorporating general purpose semi-dry and dry electrodes. Semi-dry electrodes use significantly less hydrogel that is either pre-applied or stored in an on-electrode reservoir.[6–9] These semi-dry electrodes can be more comfortable than wet electrodes and achieve similar ESIs but still require some electrolyte and can suffer from control issues such as over-release (which risks bridging). Fully dry electrodes further increase usability and patient comfort but generally result in higher ESI (>1 MΩs at <250 Hz) relative to wet-electrodes (10–100 s kΩs at <250 Hz).[10] Microneedles, pin electrodes, conductive composites, and conformal electrodes have been used to lower ESI and improve the mechanical stability of dry electrodes,[10–12] but introduce new use or fabrication trade-offs. Microneedles, which pierce the top layer of skin, achieve lower ESIs and enable higher SNR recordings. However, prolonged use of these electrodes may result in lesion formation and introduce risk of infection. Non-contact and conductive composites such as silicone carbon black and silver-glass silicone show the opposite trade-off, achieving greater comfort but suffering from higher ESIs relative to other dry electrodes.[10,11,13] Other electrode arrays have used flexible planar structures, machined metals, or metal printed devices to increase electrode compliance, comfort, and possibilities in sensing location.[14–17] Printed flexible electrode arrays enable high-density electrode placement, high-resolution 2D designs, high-volume fabrication without vacuum, and comfortable electrode compliance.
along a single axis, such as around arms.\textsuperscript{[17,18]} Recent work has focused on making chemically inert arrays out of laser-sintered gold. These 2D arrays are potentially more robust than previous demonstrations of silver and graphene-based flexible arrays but still generally require hydrogels to record through previous demonstrations of silver and graphene-based flexible 2D arrays. These 2D arrays are potentially more robust than focused on making chemically inert arrays out of laser-sintered gold. Ultimately, existing techniques emphasize that no existing single dry-electrode design can target all biopotential signals. Furthermore, existing fabrication techniques for comfortable, anatomically customized electrodes are neither low-cost nor scalable or are limited to specific electrode shapes.

Recent commercial, dry electrode ExG\textsuperscript{[23–25]} systems employ user-generic electrodes and require hairless recording locations on the wrist, forehead, or limbs to improve SNR reliability for very specific target applications across large populations. An exploratory medical device that is accepted by a patient for continuous electrophysiological recordings must be anatomically customized for a sensing location that does not interfere with daily activities. It is also preferable to record through hair without the need for shaving, contacting gels, or skin preparation without compromising SNR. To establish the range of devices that can be fabricated and that any electrode impedance would be dominated by the interface and skin itself. Detailed preparation instructions and processing for catalyst and copper plating solutions can be found below in the methods section.

To establish the range of devices that can be fabricated with this process, two dry sensor topographies were designed as examples for use in typical ExG studies. Pin electrodes with bulb tip wires were fabricated for increased patient comfort (Figure 1a) during EEG recordings through hair. Versatile circular electrodes were integrated with flexible 3D printed wristband structures for use with different arm and wrist sizes for ECG and EMG measurements (Figure 1b). When compared to clinically standard, wet electrodes of the same area, they express comparable ESIs. The results demonstrate a scalable electrode fabrication process that can rapidly produce anatomically optimized, low impedance, 3D dry electrodes for repeatable physiological recordings over long periods of time (Figure 1c). To the author’s knowledge, this is the first work to demonstrate a 3D printing and electroless plating-based fabrication process for conformal, dry biomedical electrodes.

## 2. Results

### 2.1. Fabrication Process

The electrode structures were printed using a stereolithography 3D printer (Formlabs Form 2 Printer) with a standard, clear methacrylate photopolymer (Figure 2a). Stereolithographic (SLA) printers create 3D structures by precisely laser-curing photosensitive polymer resins in a layer-by-layer fashion. This method of 3D printing results in much finer resolutions and wider choices of materials than fused deposition modeling printing, which comprises melted plastic filaments. After printing, the samples were post-processed with a 20-min IPA bath to rinse uncured resin, followed by an hour-long UV curing process to fully cure the surface.

The structures were subsequently sandblasted with 100 grit white fused aluminum oxide blasting media (Industrial Supply, Twin Falls, ID) to increase their surface area\textsuperscript{[25]} (Figure 2b). Sandblasting promotes better film adhesion and lower skin-electrode impedance. The samples were sonicated in a bath of DI water with Alconox cleaning solution for approximately 10 min before rinsing with DI water. The surface energy of the printed structures is modified in a bath of 1% benzalkonium chloride (Sigma Aldrich 12060-100G) surfactant solution for 10 min. These steps ensure the plating surface is clean and promote adhesion of the catalyst.

The metal plating process is a result of subsequent plating and cleaning steps. Prior to each plating step, the samples were rinsed and dried thoroughly. First, the 3D printed electrodes were submersed in a beaker of a palladium-tin catalyst for 10 min, followed by the copper plating solution for a minimum of 4 h, which provides a thick base layer of metal for gold plating as shown in Figure 2c. After copper plating, electrodes are soaked in the surfactant and catalyst solutions and then moved to a gold plating solution for approximately 15 min (Sigma Aldrich 901670-250ML) (Figure 2d). The second gold plating step is achieved by placing the samples in the same surfactant, catalyst, and gold plating solutions to form the top layer of the dry electrodes (Figure 2e). After all plating steps, tinned copper wires were directly soldered to the electrode surface to facilitate electrode integration with recording systems. This fabrication process ensures two 0.25 μm thick layers of gold and at least 0.5 μm of copper.\textsuperscript{[26,27]} The thickness of these two layers ensures that the plated surface acts effectively as a short circuit and that any electrode impedance would be dominated by the interface and skin itself. Detailed preparation instructions and processing for catalyst and copper plating solutions can be found below in the methods section.

### 2.2. Material Characterization

#### 2.2.1. Sheet Resistance

Sheet resistance was characterized with 4-point impedance measurements immediately after plating and once a week for nine weeks. Samples of single metal plating layers were compared against double layers, using copper as the control film. As prepared, copper-plated control samples, single layer gold...
samples, and double-layer gold samples exhibited an average sheet resistance and standard deviation of 19.2 ± 1.7, 17.6 ± 1.1, and 16.5 ± 0.7 mΩ □⁻¹, respectively. The second plated layer of gold was shown to lower the sample’s average sheet resistance as well as its standard deviation (Figure 3a) and was adopted for the longevity study. It was observed that the deposition of a single layer of gold was not adequate to prevent copper oxidation after a week. After nine weeks, an oxide layer was observed in the copper samples. In addition, a 20% increase in sheet resistance was measured. This contrasts with the 2-layer gold samples which showed no meaningful change in sheet resistance or oxide formation. (Figure 3b). The sheet resistance of both the copper and gold are displayed in reference to their respective sheet resistances on day 0.

2.2.2. Film Integrity/Acid Dip Tests

Kapton tape was applied around entire electrode surfaces and then removed. No visible gold or copper was removed with the tape. Samples also underwent nitric acid baths (Figure 3c). Nitric acid, a typical copper etchant, will readily dissolve copper but is not suitable for etching gold.²⁸ No noticeable differences were observed after dipping the samples with two gold layers.

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**Figure 1.** Additive manufacturing and electroless plating processes can rapidly fabricate dry electrodes customized for different wearables and partial limbs. This single process can produce designs ranging from dense a) pin electrodes for EEG recordings through hair to b) curved/conformal electrodes myoelectric prosthetics at scale. c) The resulting gold-finished dry electrodes can also be reused over the course of months without any skin preparation.
2.2.3. Surface Energy

3D printed surfaces require special treatments to accept durable and uniform surface platings. To improve copper adhesion to the 3D printed electrodes, surfaces were sand-blasted and treated with a surfactant. These two steps lead to an increase in surface energy as shown in Figure 3d. Water droplets on untreated surfaces exhibit contact angles of 135° while treated surfaces lead to a significantly smaller contact angle of 64°.

2.2.4. Surface Roughness

To assess the surface roughness, light microscopy photographs were taken and stylus profilometry was performed on flat, slide-like samples of the same 3D printed photopolymer (Figure 3e and Table 1). The samples were printed, masked with tape, and subjected to the same plating process described above to form selectively patterned films for comparison. A Dektak stylus profilometer was used to collect measurements on these samples at different points in the process and the surface roughness average, \( R_s \), was calculated. Due to each plating layer being \( \leq 1 \) um thick, the surface roughness was virtually unchanged after the initial sandblasting. Although it may appear that the surface roughness is unchanged before and after sandblasting, the sandblasting creates a more heterogeneous surface in all directions as opposed to the highly anisotropic structure of the cured layers of resin. From a user’s touch, and from visual inspection under optical microscopy (Figure 3e), change in surface roughness is undetectable after each plating step.

2.2.5. ESI and EDO

Impedance spectroscopy and EDO measurements with and without electrolyte gel were compared to those of gold cup electrodes. All ESI measurements were performed between two equivalent electrodes, one on the back of the head through the hair, and the other on the ipsilateral mastoid. The presented spectra in Figure 4 are half of the measurement and represent the impedance of one electrode. To emulate real-world scenarios for the dry electrodes, no skin preparation was performed before each trial and measurement sessions were repeated over the course of several weeks. Gold cup electrode measurements used typical skin cleaning prior to each trial. No skin abrasion was performed in order to limit subject discomfort. After donning the electrodes and waiting 5 min for the interface to settle, impedance measurements were performed with an LCR meter (E4980 A, Keysight). Results were fit to an equivalent circuit model using a constant phase element (CPE) (spectra shown in Figure 4a, circuit models shown in Figure 4b,c).

The dry, gold-plated, pin electrodes exhibited an average impedance of 66.7 kΩ and phase of −23° at 50 Hz. When a small amount of electrolyte gel was added to the 3D printed electrodes, the 50 Hz impedance dropped slightly to 41 kΩ while the phase increased to −15°. This is consistent with expected electrode behavior. Comparatively, clinical gold-cup electrodes (without skin abrasion) exhibited a 50 Hz impedance of 31 kΩ and phase of −24°.

Since the dry pin electrodes and wet gold cup electrodes have different contact areas, an area normalized impedance (specific impedance) should be used as a heuristic to compare their performance. The dry electrode had a contact surface area of 50 mm² and thus a specific impedance an electrode resistivity of 33 kOhmcm². The wet electrodes had a total surface area including electrolyte gel of roughly 70 mm² and thus exhibits a specific impedance of 21.7 kOhmcm². Even without skin abrasion, the wet electrode still has a lower resistivity overall, but the dry electrode performance is significantly more area efficient than existing wearable dry electrodes. Furthermore, the dry electrode’s gold finish was stable over time. No noticeable surface degradation was observed despite daily reuse and cleaning over the course of two months nor did the measured impedance values increase throughout the observation period (Table 2).

EDO measurements were taken between each dry sensing electrode and an equivalently sized dry reference electrode with the WAND mini wireless recording device.
(see Methods section) with an input range up to 400 mV and an input impedance of 10 MΩ. No skin cleaning or abrasion was performed. Initially, the mean EDO and standard deviation was $-40 \text{ mV} \pm 15 \text{ mV}$ ($n = 10$). After approximately 20 min, the EDO settled to $-20 \text{ mV} \pm 10 \text{ mV}$ ($n = 10$). The minimum and maximum EDO values after settling are $-32$ and $23 \text{ mV}$, respectively. It is important to note that the measured EDO is not equivalent to the electrode’s open-circuit potential due to the recording front end’s finite input impedance.

2.3. ExG Measurements

ECG, EMG, and EEG signals were recorded using the WAND mini system (see Methods for more details). Three 21–25 year old subjects (one male, two female) were recruited to perform proof-of-concept ExG measurements. To mimic realistic day-to-day scenarios, no skin preparation was performed prior to dry electrode donning in all experiments. When wet electrodes were used for comparison measurements, a trained technician performed skin cleaning, hydrogel application, and electrode
placement. It is important to note that pressure behind electrodes can result in lower ESI and improve recorded signal quality, but it is unrealistic to expect users to apply great pressure on wearables used on a daily basis. Thus, subjects were tasked with donning the wearable dry electrodes themselves and adjusting the bands until the electrodes were comfortably in contact with their skin. The proof-of-concept study was approved by UC Berkeley’s Institutional Review Board (CPHS protocol ID: 2018-09-11395).

2.3.1. Electrocardiography (ECG)

Cross-body ECG measurements were performed using 3-electrode armbands with embedded curved electrodes on each arm. Differential measurements were taken across both arm bands to record a cross-body ECG (Figure 5a). One of the dry electrodes (Figure 5b) was selected as a ground to reduce interference in the recordings. A clear ECG rhythm (PQRST complex) was recorded and is labeled in Figure 5a.

2.3.2. Electromyography (EMG)

Bicep EMG was recorded on a single subject wearing two bands on a single-arm, one across the bicep and another by the elbow. The elbow electrodes were used as both a system ground and as references for the bicep electrodes. Sensing electrodes were placed along the bicep. Users were cued to flex their biceps every 5 seconds, resulting in clear increase in broad spectrum (0–200 Hz) electrical activity consistent with large scale muscle activity both in frequency and time domain (Figure 5c,d). The electrode arrangement is shown in Figure 5e.

2.3.3. Electroencephalography (EEG)

A single user’s alpha attenuation response was measured with both wet and dry electrode-based recording setups. Alpha rhythms are a spontaneous neural signal centered between 8–12 Hz that reflects a person’s state of attention and can be modulated by opening and closing their eyes. Originating from the occipital lobe, alpha waves are large amplitude signals that are commonly used to benchmark EEG systems. To make a comparison between EEG recorded with dry and wet electrodes, back-to-back alpha attenuation measurements were performed with the same subject, recording sites, and instrumentation. First, the user would wear a 3D printed headband with pin-style dry electrodes. The headband was oriented to roughly correspond to T3 and T4 recording sides (according to the traditional 10–20 map). Afterward, the dry electrodes were replaced with clinically standard gold cup electrodes (Figure 4b) and the experiment was repeated. To reduce interference and make the comparison as consistent as possible between the two measurements, a single wet ground electrode was placed on the subject’s left mastoid and maintained for both wet and dry electrode measurements. In both setups, the subject was tasked to close/open their eyes every 30 s to modulate alpha band activity. When the subject closed their eyes, alpha band power increased while ocular artifacts decreased (Figure 5f). The results showed comparable performance between the wet and dry electrodes.

Table 1. Sample surface roughness at different points along the plating procedure.

<table>
<thead>
<tr>
<th>Bare print</th>
<th>Sandblasted surface</th>
<th>Cu 1 layer Au</th>
<th>2 layer Au</th>
</tr>
</thead>
<tbody>
<tr>
<td>Avg Roughness, $R_a (\mu m)$</td>
<td>$3.34 \pm 0.90$</td>
<td>$3.34 \pm 0.67$</td>
<td>$4.32 \pm 1.10$</td>
</tr>
</tbody>
</table>

Table 2. Au Plated Pin Electrodes electrode–skin interface impedance over 60 days. Every 30 days the average ($n = 5$) ESI magnitude and phase are measured. The recording site is neither cleaned nor abraded before measurements. Day-to-day variation consistent with [13,4] is observed but is likely due to skin condition given visual consistency of electrode coatings.

<table>
<thead>
<tr>
<th>Time [Days]</th>
<th>0</th>
<th>30</th>
<th>60</th>
</tr>
</thead>
<tbody>
<tr>
<td>50 Hz impedance magnitude</td>
<td>86.3 kΩ</td>
<td>50.1 kΩ</td>
<td>63.7 kΩ</td>
</tr>
<tr>
<td>50 Hz impedance phase</td>
<td>$-28^\circ$</td>
<td>$-18^\circ$</td>
<td>$-23^\circ$</td>
</tr>
</tbody>
</table>

Figure 4. Electrode–skin interface (ESI) Characterization of Au Plated Pin Electrodes vs Clinical Au Cup Electrodes on across two months. a) Average ($n = 15$) magnitude and phase of gold-plated pin electrodes on scalp with and without electrolyte gel alongside gold cup electrodes. An electrode model with a CPE model, $Z_{elec}$ is fitted to the dry gold-plated pin electrodes. b) 3D-printed, gold-plated, dry scalp electrode, and clinical gold cup electrode. c) CPE model with fitted parameters for spread resistance $R_S$, charge transfer resistance $R_{CT}$, and CPE double layer, $CPE_{DL}$.
Figure 5. Sample physiological measurements taken with gold-plated, dry electrodes. a) Time domain measurement \((n = 1)\) of single lead, cross-body ECG measurement with labeled PQRST complexes. b) ECG measurement set up with 3D-printed bands with dry electrodes on each arm. c) Spectrogram and d) time-domain of EMG measurements \((n = 1)\) taken with e) two electrode bands placed along a single subject’s arm. The subject is cued to flex their bicep every 5 s. Characteristic broad spectrum muscle activity \((0–200 \text{ Hz})\) is observed when the user is cued to flex. f) Sample Time-frequency spectrogram of a single dry electrode alpha attenuation response. g) Averaged \((n = 3)\) alpha \((8–12 \text{ Hz})\) band power of alpha modulation recorded across a subject’s scalp for both dry pin and wet gold cup electrode measurements. Alpha power increased by a factor of \(5 \times\) in the eyes-closed state for both dry and wet measurements. h) Alpha attenuation response measurement setup. Two dry electrodes are placed on either side of the head.
Table 3. User-specific scoring of the dry electrode headband, dry electrode arm band, and wet electrodes. Dry electrodes are rated significantly more comfortable due to the lack of skin preparation and hydrogel.

<table>
<thead>
<tr>
<th>Electrode</th>
<th>User 1</th>
<th>User 2</th>
<th>User 3</th>
<th>Average</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pin Dry Electrode</td>
<td>4</td>
<td>4</td>
<td>4</td>
<td>4</td>
</tr>
<tr>
<td>Curved Dry Electrode</td>
<td>5</td>
<td>4</td>
<td>4</td>
<td>4.3</td>
</tr>
<tr>
<td>Wet Electrode</td>
<td>3</td>
<td>2</td>
<td>2</td>
<td>2.3</td>
</tr>
</tbody>
</table>

Both wet and dry electrode setups recorded alpha modulation ratios of 4.89 and 4.91, respectively (Figure 5g).

2.3.4. Dry Electrode Comfort

Subjects donned the curved, dry electrode armband (Figure 5e), dry pin-electrode headband (Figure 5h), and clinically standard, wet, gold cup electrodes (Figure 4b) for three 2 h sessions. After the third session, subjects asked to rate the comfort of each electrode form factor on a scale of 1–5 (1: unbearably uncomfortable and 5: extremely comfortable). User scores are in Table 3. Wet electrodes had an average rating of 2.3 with subjects citing skin abrasion and residual gel as defining uncomfortable factors. Both dry electrodes were rated 4 on average. The dry electrodes had no skin preparation and left no lingering skin irritation, soreness, or redness. Subjects commented that the dry electrodes felt like wearing jewelry and could be worn long term. Wet electrode sites exhibited minor irritation and lingering redness.

3. Discussion

This multi-layer electroless plating fabrication technique enables the rapid prototyping of customized electrodes for any application. Unlike techniques such as sputtering, evaporation, or spin-coating, the presented electroless plating process will evenly metalize any electrode shape with a gold finish, enabling optimization for any recording location without vacuum or constraints on surface feature density, overhangs, and assembly. The resulting electrode surfaces are robust enough to be directly soldered to, which allows integration with the WAND mini and other systems that use conventional sensor data processing and transmission electronics. Further, the WAND mini and other systems that use conventional vacuum or constraints on surface feature density, overhangs, enabling optimization for any recording location without vacuum or constraints on surface feature density, overhangs, and assembly. The WECG, EMG, and EEG experiments comprehensively demonstrate the potential of this optimized fabrication process in sample use cases. In ECG, clear waveforms were shown, demonstrating potential to detect heart rate, heart rate variability, and cardiac arrhythmias. The presented EMG exhibited expected broadband muscle activity. With EEG, the alpha attenuation response recorded with the dry electrodes exhibited the same alpha modulation as the wet electrode case. All three signal paradigms were recorded without any skin cleaning, abrasion, or hydrogel.

4. Experimental Section

Plating Solutions: Both the catalyst and copper plating solutions were made in house. The catalyst solution was prepared between 60–70 °C and was stirred for approximately 1 h after all components were added. This solution was prepared in full and lasts several weeks before the salts precipitate and were no longer usable.

The two main components for an electroless plating solution were a metallic salt and a reducing agent, in this case, copper(II) sulfate and formaldehyde respectively. At a sufficiently high pH (the solution was adjusted to 12.8 by adding NaOH), formaldehyde reacts with hydroxide ions in solution to reduce copper ions in the salt:

$$\text{Cu}^{2+} + 2\text{HCHO} + 4\text{OH}^- \rightarrow \text{Cu} + \text{H}_2\text{O} + 2\text{H}_2\text{O} + 2\text{HCO}_3^- \tag{1}$$

In the solution, EDTA was added, which acts as a complexing agent (as copper salts were insoluble at pH > 4), and ferrocyanide, which acts to stabilize the solution over time (the solution was stored without the addition of formaldehyde, which was then added to the appropriate quantity being used for plating). Layers were typically built at a rate of about <1 µm h⁻¹ at ambient temperature, so leaving the samples...
for several hours or overnight in a covered plating solution provides sufficient coverage. A lightly bubbling nitrogen line was left in the solution to provide light agitation, promote even coverage by displacing the hydrogen gas product, and limit oxidization of the copper during the plating process.

Lastly, the gold layer was applied by heating the gold plating solution to about 90 °C and submerging the samples for about 15 min. This process was self-limiting, as gold layers adhere to the copper only and not upon itself. Solution recipes can be found in Table 4.

Experimental ExG Recording System: In this work, ExG signals were acquired using a miniature, wireless, artifact-free neuromodulation device (WAND mini), a low-profile, custom neural recording system that streams recorded data over Bluetooth low energy (BLE) to a base station connected to a laptop (Figure 6). WAND mini was derived from a previous design for a wireless, artifact-free neuromodulation device (WAND), reduced to a form factor of 2.5 × 2.5 cm², and embedded with custom firmware (Figure 6). Recording and digitization were performed by a custom neuromodulation IC (NMIC, Cortera Neurotechnologies, Inc.) integrated with 64 digitizing frontends, thereby expandable to recording applications with higher electrode counts. NMIC and WAND mini specifications are listed in Table 5.

The NMIC was selected for its low power and high dynamic range, supporting a 100–400 mV input range with a flat input-referred noise voltage spectrum of 70 nV √ Hz⁻¹. The analog-to-digital converters (ADCs) have a resolution of 15 bits and sample at 1 kSps, providing sufficient resolution and bandwidth for EMG, ECG, and EEG signals. The wide linear input range can accommodate the large EDOs and provides robustness to interference.

Analytical Methods: All data presented as mean and standard deviations assume normally distributed data sets with equal variances. For physiological data, minimal pre-processing and normalization were performed. All presented physiological measurements (ECG, EMG, and EEG) were filtered with 60 Hz, 120 Hz, and 180 Hz notch filters using MATLAB. No normalization was performed. ECG in Figure 5a was centered around 0 V via mean removal in MATLAB.

Supporting Information
Supporting Information is available from the Wiley Online Library or from the author.

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Conflict of Interest
The authors declare no conflict of interest.

Data Availability Statement
The data that support the findings of this study are available on request from the corresponding author. The data are not publicly available due to privacy or ethical restrictions.

Table 4. Plating process solution components and purpose. All materials are purchased from Sigma-Aldrich.

<table>
<thead>
<tr>
<th>Solution</th>
<th>Components</th>
<th>Purpose</th>
</tr>
</thead>
<tbody>
<tr>
<td>Catalyst</td>
<td>1 L deionized water</td>
<td>Provides very thin initial palladium layer for copper adhesion</td>
</tr>
<tr>
<td></td>
<td>60 mL HCl</td>
<td></td>
</tr>
<tr>
<td></td>
<td>0.25 g PdCl₂ 12 g SnCl₂, after PdCl₂ completely dissolves</td>
<td></td>
</tr>
<tr>
<td>Electroless copper plating</td>
<td>1000 mL deionized water</td>
<td>Plates a thick layer of highly conductive material</td>
</tr>
<tr>
<td></td>
<td>18 g CuSO₄·5H₂O</td>
<td></td>
</tr>
<tr>
<td></td>
<td>48 g EDTA</td>
<td></td>
</tr>
<tr>
<td></td>
<td>57.2 mg K₄Fe(CN)₆·3H₂O</td>
<td></td>
</tr>
<tr>
<td></td>
<td>1 mL HCl</td>
<td></td>
</tr>
<tr>
<td></td>
<td>NaOH as needed to adjust pH to 12.8</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Formaldehyde, when ready for use in 22.5:1 ratio of plating solution/formaldehyde</td>
<td></td>
</tr>
<tr>
<td>Electroless gold plating</td>
<td>Bright electroless gold plating solution (Sigma-Aldrich Part Number: 901670)</td>
<td>Prevents copper oxidation and improves biocompatibility</td>
</tr>
</tbody>
</table>

Table 5. Relevant NMIC and WAND mini specifications to for presented ExG measurements.

<table>
<thead>
<tr>
<th>NMIC and WAND mini Specifications</th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>Max Recording channels</td>
<td>64</td>
</tr>
<tr>
<td>Input range</td>
<td>100–400 mV</td>
</tr>
<tr>
<td>Input referred noise voltage spectrum</td>
<td>70 nV √ Hz⁻¹</td>
</tr>
<tr>
<td>Input impedance</td>
<td>40 MΩ</td>
</tr>
<tr>
<td>ADC resolution</td>
<td>15 bits</td>
</tr>
<tr>
<td>ADC sample rate</td>
<td>1 kS s⁻¹</td>
</tr>
<tr>
<td>Wireless data rate</td>
<td>2 Mbps</td>
</tr>
<tr>
<td>Board dimensions</td>
<td>25.4 mm × 25.4 mm</td>
</tr>
</tbody>
</table>

Figure 6. WAND mini neural recording module.
Keywords
3D printing, electrocardiography, electroencephalography, electroless plating, electrophysiological sensors, medical devices, prosthetics

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